# Plant Physiology and Genetics

**Module “Applied Genetcis”**

Prof. Lorenzo Stagnati

***COURSE AIM AND INTENDED LEARNING OUTCOMES***

The module is carried out in the 2nd term and consists of 4.0 CFU (32 hours) of lectures and 1.0 CFU (12 hours) of practical work. The module is divided in two parts consisting of 2.0 CFU. The first part aims to: 1) illustrate the most basics knowledge on genetics; 2) explain the most important biological processes involving DNA and RNA (DNA replication, transcription and translation). The second part aims to present, in the form of seminars, the most important practical applications of DNA based knowledge and technologies to issues as agriculture sustainability and food traceability.

Learning outcomes:

After successful completion of the module, students are expected to be able:

- to have the most important basic knowledge concerning DNA, the biological processes involving it and to describe them with a special focus on 1) the flow of the genetic information from the DNA to the proteins, 2) DNA replication, transcription ad translation, 3) the structure of DNA and chromosomes in both prokaryotic and eukaryotic organisms.

- to explain the different kinds of mutations, the mechanisms inducing mutations and the effects of mutations on the genetic code and on the phenotype of the organisms;

- to understand and to explain the use of DNA for the characterisation and preservation of plant biodiversity;

- to understand what Genetically Modified Organisms are, how GMOs can be produced and their importance for modern agriculture sustainability;

- to describe the use of DNA markers for food traceability;

- to carry out a DNA extraction and a PCR reaction for analysing plant biodiversity.

***COURSE CONTENT***

Applied Genetics

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| --- | --- |
|  | CFU |
| **First part – Basics on genetics** | **2.5** |
| DNA and the genome – composition, structure, organisation in chromosomes (nuclear and organellar chromosomes), DNA replication, viral, prokariotic and eukaryotic genomes, basics on Mendelian genetics | 1.25 |
| The transcription – the genetic code, the RNA, different kinds of RNAs, prokariotic and eukaryotic transcription; | 0.4 |
| The translation, from RNA to proteins; | 0.4 |
| Mutations – different kinds of mutations, mechanisms producing mutations, genetic and phenotypic consequences of mutations. | 0.45 |
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| **Second part – Practical applications of genetics and DNA knowledge** | **1.5** |
| PCR (Polymerase Chain Reaction) | 0.25 |
| DNA based traceability of plant based productions | 0.25 |
| Germplasm management, *Ex situ* and *In situ* preservation of plant biodiversity | 0.25 |
| DNA based markers development, determination of the genetic fingerprint | 0.25 |
| Transgenesis, cisgenesis and genome editing | 0.5 |
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| **Practical work** | **1.0** |
| Plant DNA extraction; evaluation and quantification of nucleic acids, PCR (Polymerase Chain Reaction), | 0.75 |
| Application of Mendelian genetics | 0.25 |

***READING LIST***

W.S. KLUG, M.R. CUMMINGS, C.A. SPENCER, PALLADINO M.A. Essential of Genetics, Pearson, 2017.

Papers from the scientific literature that will be provided by the teacher.

Power point presentations will be made available during the course before the beginning of each new topic.

***TEACHING METHOD***

The teaching method will embrace the following activities:

1) Indoor class where main course topics will be covered along with several applied examples. Each new lesson, starting from the second one, will begin with a 10 – 15 minutes refresher of the previous one to recall the main subjects previously addressed. Questions will be posed to stimulate discussion.

2) Laboratory practical activities and exercises aimed at understanding the use of DNA analysis to evaluate plant biodiversity. The different laboratory sessions will cover DNA extraction and evaluation, PCR analysis and evaluation of the results. Applications of Mendelian genetics will focus on the resolution of problems covering the main aspects of mendelism.

***ASSESSMENT METHOD AND CRITERIA***

The assessment will take the form of two written tests consisting of 16 questions each one regarding the main arguments of the course, lasting 45 minutes. They will consist of multiple-choice questions (1 point for each correct answer, 0 point for each wrong answer) with students choosing from four possible answers. Pass mark for both the tests is 18/30 out of 30/30. A bonus mark (lode) will be awarded to those students who will correctly answer to all the questions. The final score of the module will be the average of the scores of the two tests. The first test will take place at the end of the course and it will be focused on DNA and the genome. In case of positive results in the first test, the second written test, that will take place at the official exam dates, will be focused on Transcription, Translation and Mutations. In case of negative results in the first test, or for students unwilling to take the first test, the assessment will take the form of a single written test consisting of 32 multiple choice questions, lasting 90 minutes, on the main arguments of the course to be held at the official exam dates.

The final mark out of thirty of the “Applied Genetics” module will be further averaged out (to give a mark out of thirty) with the mark obtained in the test of the “Plant physiology” module to obtain the final mark of the course Plant Physiology and Genetics.

***NOTES AND PREREQUISITES***

The course does not need particular prerequisites.

Prof. Matteo Busconi is available to meet with students every day at the Department of Sustainable Crop Production - Agronomy and Plant Biotechnology Area, or online using the platform Microsoft teams.

**Module “Plant Physiology”**

Prof. Alessandra Lanubile

***COURSE AIMS AND INTENDED LEARNING OUTCOMES***

The course is carried out in the 2nd term and consists of 3.5 CFU (28 hours) of lectures and 0.5 CFU (6 hours) of practical work. The course aims to: 1) illustrate the characteristics of plant cells; 2) describe the main features of primary and secondary tissues; 3) explain the anatomy of organs as leaves, stem and root.

Learning outcomes:

After successful completion of this course students are expected to be able to:

- explain the main structures of plant cells, more in detail, the components of primary and secondary cell wall, vacuole and plastids;

- describe the characteristics and the organization of different cell types that form meristematic, dermal, parenchymatic, mechanical, transport and secretory tissues;

- explain the primary and secondary structure of stem and root, the anatomy of leaves, highlighting the main differences between the different groups of plants;

- explain the activities of meristematic tissues, in particular how they differentiate into other types of tissues as they mature to generate adult tissues and organs;

- to identify through optical microscope observation the primary and secondary stem of Angiosperm and Gymnosperm plants, to distinguish dorsiventral and isobilateral leaves, to recognize Angiosperm mocot and dicot roots.

***COURSE CONTENT***

Plant Physiology

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|  | CFU |
| **Cellular level**  The plant cell, the cell wall, the endomembrane system, the vacuole, the plastids, the nucleus. | **1.0** |
| **Tissue level**  Primary and secondary meristematic tissues, dermal, parenchymatic, mechanical, transport and secretory tissues. | **1.0** |
| **Organ level**  The stem: primary body, formation of the fascicular and cork cambium, secondary body.  The leaf: structure and arrangement, anatomy, modifications.  The root: zonation pattern, anatomy, adaptations. | **1.0** |
| Plant reproduction systems, differentiation of stem, leave, root, flowers, seed and fruit. | **0.5** |
| **Practical work** Optical microscope observation of the tissues and the primary and secondary body of roots, stems and leaves. | **0.5** |

***READING LIST***

R.F. EVERT, *Esau’s Plant Anatomy*, Wiley, Hoboken (new jersey), 2006

D.F. Cutler, T. Botha, D.W. Stevenson, *Plant Anatomy: an applied approach*, Blackwell Publishing, Ma (USA), 2008

Power point presentations will be made available during the course.

***TEACHING METHOD***

The teaching method will embrace the following activities:

1) Indoor classes where main course topics will be covered along with several applied examples.

2) Laboratory activities aimed at optical microscope observations of anatomical preparations of primary and secondary stem of Angiosperm and Gymnosperm plants, dorsiventral and isobilateral leaves, Angiosperm mocot and dicot roots. The frequency of practical work is mandatory.

***ASSESSMENT METHOD AND CRITERIA***

The assessment will take the form of a written test consisting of one open question (maximum score of 14 points) and the optical microscope observation and detailed description of two histology slides of roots, stems and leaves (maximum score of 8 points, each). The test lasts 60 minutes. Pass mark is 18/30 out of 30/30.

***NOTES AND PREREQUISITES***

The course does not need particular prerequisites.

Prof. Alessandra Lanubile is available to meet students after class at the Department of Sustainable Crop Production - Agronomy and Plant Biotechnology Area.

In case the current Covid-19 health emergency does not allow frontal teaching, remote teaching will be carried out through synchronous or asynchronous procedures that will be promptly notified to students

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Prof. Matteo Busconi

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| DNA based traceability of plant based productions | 0.25 |
| Germplasm management, *Ex situ* and *In situ* preservation of plant biodiversity | 0.25 |
| DNA based markers development, determination of the genetic fingerprint | 0.25 |
| Transgenesis, cisgenesis and genome editing | 0.5 |
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| **Practical work** | **1.0** |
| Plant DNA extraction; evaluation and quantification of nucleic acids, PCR (Polymerase Chain Reaction) | 1.0 |

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